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# REVIEW



# Impacts of artificial barriers on the connectivity and dispersal of vascular macrophytes in rivers: A critical review

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# **Abstract**

- 1. Macrophytes play important functional roles in river ecosystems, providing habitat and food, as well as influencing flow, water chemistry, and sediment dynamics. They also represent an important component of river biodiversity.
- 2. Artificial river barriers have the potential to disrupt macrophyte dispersal, and compromise their distribution and persistence, but little information is available compared to barrier impacts on fish and macroinvertebrates. Here, we review the mechanisms supporting dispersal of river macrophytes in rivers and evaluate the nature of barrier impacts on macrophytes.
- 3. Hydrochory (dispersal of propagules by water) is the principal mechanism of downstream dispersal, while zoochory (dispersal of propagules by animals) is likely to be the most important vector of upstream dispersal and inter-catchment transport.
- 4. Most studies have focused on the impact of large structures such as dams, and the findings indicate the impact is highly context dependent. Slow-flowing habitats upstream of dams can act as traps to drifting propagules and thereby interrupt hydrochory. However, the consequences of interrupted hydrochory for downstream populations are unclear. River regulation can result in lower macrophyte diversity, although the lentic habitats associated with reservoirs can also favour an increase in the abundance and richness of macrophyte communities.
- 5. Instream barriers are unlikely to affect zoochory by birds directly, but barriers are well known to restrict fish movements, so there is considerable potential for barriers to disrupt zoochory by fish, although no empirical study has specifically examined this possibility.
- 6. There is a paucity of studies examining the impacts of low-head barriers on macrophyte dispersal. Given the influence of macrophytes on river processes, we call for further research into barrier impacts on macrophyte population dynamics in order to gain a better understanding of the consequences of river fragmentation for fluvial communities and ecosystem functioning.

# KEYWORDS

aquatic plant, dams, hydrochory, river regulation, zoochory

# 1 | INTRODUCTION

Aquatic vascular macrophytes (Tracheophyta) are an important component of river biodiversity, including over 2,600 species from 88 different families globally (Chambers, Lacoul, Murphy, & Thomaz, 2008). Apart from their contribution to aquatic biodiversity, macrophytes play important functional roles in river ecosystems: they provide food resources and habitat (Biggs, 1996; Grenouillet, Pont, & Seip, 2002), and act as ecosystem engineers by trapping sediments and altering flow dynamics (Carpenter & Lodge, 1986; Gurnell, Oosterhout, Vlieger, & Goodson, 2006; Horvath, 2004). Macrophytes can also regulate water chemistry (Clarke & Wharton, 2001), and are one of the key metrics used to measure the ecological status of river systems under the Water Framework Directive (European Commission, 2000). Hence, maintaining healthy macrophyte communities is essential for river ecosystem function.

Artificial barriers, such as dams and weirs, can have a pervasive influence on river systems (Ellis & Jones, 2013). However, while barrier impacts on fish populations have received considerable attention (Fullerton et al., 2010; Morita & Yamamoto, 2002; Perkin & Gido, 2012), impacts on aquatic macrophytes remain relatively unexplored. Connectivity is essential for the resilience of freshwater biota and the maintenance of river processes (Fagan, 2002; Pringle, 2001, 2003). Artificial barriers alter river hydrology and create discontinuities in substrate composition, temperature regime, and water chemistry (Mueller, Pander, & Geist, 2011) that could potentially disrupt macrophyte dispersal and population structure. Furthermore, river obstacles affect the distribution and movements of fish (Branco, Amaral, Ferreira, & Santos, 2017; Dehais, Eudeline, Berrebi, & Argillier, 2010; Diebel, Fedora, Cogswell, & O'Hanley, 2015), as well as birds indirectly by affecting habitat availability (Nilsson & Dynesius, 1994; Stevens, Buck, Brown, & Kline, 1997), and these groups can act as important dispersal vectors for riverine macrophytes (Charalambidou & Santamaría, 2002; Horn, 1997; Pollux et al., 2006). There are estimated to be over 16 million barriers in river systems worldwide (Lehner et al., 2011), and recent studies suggest that even this number is likely to be a severe underestimate because the abundance of low-head barriers is not well known (Garcia de Leaniz et al., 2018; Jones et al., 2019). Hence, there is considerable potential for barriers to have wide-reaching effects on macrophyte abundance and distribution.

Invasive species are one of the leading causes of decline in freshwater biodiversity worldwide (Dudgeon et al., 2006; Reid et al., 2019), and the effects of non-native macrophytes can be particularly damaging because they can exclude native species, alter habitat complexity, disrupt food webs, modify sediment dynamics, cause hypoxia, release allelopathic chemicals, and facilitate the establishment of other exotic species (Bunn, Davies, Kellaway, & Prosser, 1998; Fleming & Dibble, 2015; Michelan, Thomaz, Mormul, & Carvalho, 2010; Schultz & Dibble, 2012). Invasive macrophytes generally have higher growth rates, higher plasticity, and disperse more readily than native species (Schultz & Dibble, 2012; Umetsu, Evangelista, & Thomaz, 2012), traits that may make them more

suited to establishment in regulated rivers. Hence, it is important to consider the effect of river barriers on invasive macrophytes.

Several studies have examined macrophyte dispersal and settlement dynamics in rivers (Anderson, Nuttle, Saldaña Rojas, Pendergast, & Flecker, 2011; Brochet, Guillemain, Fritz, Gauthier-Clerc, & Green, 2010; Johansson & Nilsson, 1993; Pollux, 2011; Riis, 2008), and the impact of large barriers has been investigated in a number of case studies (Ceschin, Tombolini, Abati, & Zuccarello, 2015; Merritt & Wohl, 2006; Vukov et al., 2018). However, barrier impacts on macrophyte dispersal have not been reviewed. Here, we review the current literature on macrophyte dispersal and settlement, and examine the impacts of anthropogenic barriers on population dynamics.

# 2 | METHODS

We searched for relevant literature using Google Scholar and Web of Science search engines, utilising different combinations of search terms depending on the subsection of the review (see Table S1 for glossary of terms and Table S2 for search terms and number of hits). No restrictions on year of publication or type of document were imposed. As not all search results were pertinent to the scope of the review, they were systematically reviewed by the lead author. For example, for the section on hydrochory, publications were deemed relevant only if they focussed on aquatic plants, related to fluvial systems, and dispersal by water was the focus of the paper. The use of search term strings sometimes returned very large numbers of hits (see Table S2) so results were ordered in terms of relevance (i.e. records containing highest number of keywords first), and the first 200 records were reviewed for relevance by the lead author. As predefined strings of search terms may not always be effective in returning the most relevant material, key publications highlighted during reading were also added to the reference list. By following the steps detailed above we are confident that we effectively covered the most important literature on the topic.

# 3 | MACROPHYTE DISPERSAL AND SETTLEMENT

Fluvial ecosystems are inherently dynamic environments (Ward & Stanford, 1995), exposed to regular flow disturbances that cause local extirpations of macrophyte communities (Franklin, Dunbar, & Whitehead, 2008; Riis & Biggs, 2003). Aquatic plants have various adaptations that facilitate dispersal, and allow them to recolonise vacant habitat patches (Catford & Jansson, 2014). Longitudinal dispersal in river networks is also important for maintaining genetic diversity of populations (Honnay, Jacquemyn, Nackaerts, Breyne, & Looy, 2010; Horreo et al., 2011; Pollux, Santamaria, & Ouborg, 2005). Dispersal of propagules (including whole plants, vegetative parts, and seeds; Thomaz, Mormul, & Michelan, 2015) can take place via one of four mechanisms (Figure 1): river flow (hydrochory),

Anemochory - wind

dispersal of seeds.

apart from during

Multidirectional.

Settlement and colonisation in slow flowing, shallow areas

Emergent taxa with

small light seeds only.

Short distance dispersal

extreme weather events.

Non-buoyant propagules can be transported in the bedload

Thick macrophyte beds trap

drifting propagules

Hydrochory – dispersal of propagules in river flow. Mainly plant fragments, but also seeds and seedlings. Dispersal distance depends on flows and speciesspecific traits. Dominant mechanism of downstream dispersal. Unidirectional.

Anthropochory – dispersal by human movement. Can spread invasive species. Role in movement of native species unexplored. Multidirectional.

Birds and humans can move propagules between catchments

FIGURE 1 The main mechanisms driving dispersal and settlement of macrophytes in rivers

movement by animals (zoochory), by wind (anemochory), or human-mediated dispersal (anthropochory).

# 3.1 | Hydrochory

Hydrochory is considered to be a principal dispersal vector for macrophytes in free-flowing rivers (Dawson, 1988; Merritt & Wohl, 2002). In contrast to terrestrial plants, many macrophytes disperse largely through vegetative parts such as rhizomes, stolons, tubers, turions, stem fragments, and even entire plants (Boedeltje, Bakker, Brinke, Groenendael, & Soesbergen, 2004; Sand-Jensen, Andersen, & Andersen, 1999; Umetsu et al., 2012). Aquatic macrophytes often have functional adaptations to facilitate dispersal in flows (Catford & Jansson, 2014), including air-filled structures and hairs that trap air bubbles to increase buoyancy, and therefore expedite drift (Riis & Sand-Jensen, 2006).

Production of vegetative fragments occurs either through breakage caused by water currents or animal disturbance (allofragmentation; see Madsen, Eichler, & Boylen, 1988), or via autofragmentation—the release of tissue as a plant reaches peak biomass (Riis, Madsen, & Sennels, 2009). Species with more streamlined morphological adaptations (e.g. *Ranunculus* spp.) are less susceptible

to stem breakage and uprooting by flows compared to those with higher hydraulic resistance and weaker rooting strength (e.g. *Rorippa nasturtium aquaticum*; Sand-Jensen, 2003). The structural properties of the vegetative bodies also affect how far they drift. For instance, Riis and Sand-Jensen (2006) found that the majority of denser *Elodea canadensis* fragments settled within 0.3 km of the source plant, whereas *Ranunculus peltatus* stems, which were more buoyant, tended to disperse longer distances (up to 5 km). These observations reflect the importance of species-specific traits in determining dispersal distance in river flows (Catford & Jansson, 2014).

In emergent taxa and species associated with river margins, seed dispersal can be more important, and experimental evidence suggests that hydrochorous seed dispersal is a major mechanism structuring plant communities along rivers (Nilsson, Ekblad, Gardfjell, & Carlberg, 1991; Nilsson, Gardfjell, & Grelsson, 1991; Pollux, Luteijn, Groenendael, & Ouborg, 2009). Many species have buoyant seeds to expedite dispersal (Nilsson, Brown, Jansson, & Merritt, 2010), with dispersal rates of up to 15 km/hr recorded in the floating seeds of *Polygonum* sp. (Staniforth & Cavers, 1976). The seeds of some other aquatic plants (e.g. *Juncus* spp.) are negatively buoyant and sink immediately after being liberated, but the young seedlings float and can travel large distances before taking root (Barrat-Segretain, 1996). Even dense seeds can be transported large distances in the

bedload of rivers (Markwith & Leigh, 2008, 2012) or on floating debris rafts (Skoglund, 1989).

Dispersal and settlement dynamics are highly dependent on hydrology. Many species time the release of propagules to coincide with high flows to facilitate long-distance dispersal (Catford & Jansson, 2014). High water velocities increase drag on drifting plant fragments and, therefore, reduce the likelihood of settlement (Sand-Jensen, 2003). Fast flows also compress plant growth against the stream bed, reducing roughness, and thereby reduce retention rates of drifting propagules (Sand-Jensen, 2003). Hence, high water velocities favour long-distance dispersal, while slow flows, often associated with meanders, are required for propagule settlement. Dispersal distance tends to be positively associated with the width of the stream channel, drift often being higher in large rivers than in smaller streams (Riis & Sand-Jensen, 2006). Stem fragments tend to be more frequently deposited in shallower areas of river channels (Riis & Sand-Jensen, 2006), and in areas with high bed roughness, or where there is thick vegetative growth (Riis, 2008).

It is widely accepted that hydrochory is the dominant mode of downstream dispersal in river macrophytes, and results in effective dispersal across scales ranging from hundreds of metres to kilometres (Boedeltje et al., 2004; Nilsson, Ekblad, et al., 1991; Nilsson, Gardfjell, et al., 1991; Riis & Sand-Jensen, 2006). Hence, hydrochorous dispersal is critical to the maintenance of genetic diversity in macrophyte populations. A number of studies have found that the unidirectional flow of water results in asymmetrical gene flow (Gornall, Hollingsworth, & Preston, 1998; Pollux et al., 2009). For instance, Pollux et al. (2009) found a significant increase in the genetic diversity of Sparganium emersum populations with distance downstream, gene flow being approximately 3.5 times higher in a downstream direction than upstream. In the absence of a mechanism for upstream dispersal, the continual downstream drift of propagules via hydrochory would theoretically result in loss of genetic diversity, and eventually population collapse of macrophytes in headwaters (Honnay et al., 2010; Pollux et al., 2009). However, many studies have found no evidence of genetic impoverishment in upstream populations (Chen, Li, Yin, Cheng, & Li, 2009; Honnay et al., 2010; Markwith & Scanlon, 2007; Tero, Aspi, Siikamäki, Jäkäläniemi, & Tuomi, 2003). It is largely unknown why some populations show evidence of upstream genetic impoverishment while others do not (see Honnay et al., 2010; Markwith & Scanlon, 2007; Tero et al., 2003) but the fact that macrophyte populations can persist in headwaters is good evidence that vectors for upstream dispersal must exist.

# 3.2 | Zoochory

Zoochory (movement by animals) plays an important role in longitudinal movements of plant propagules along rivers, and has been demonstrated in fish (Pollux et al., 2006), birds (Figuerola & Green, 2002), mammals (Medwecka-Kornaś & Hawro, 1993), and reptiles

(Padgett, Carboni, & Schepis, 2010). Transport of propagules can either take place inside the gut of animals (endozoochory), or attached to their bodies (ectozoochory).

A wide range of fish species are known to consume plant seeds (Correa, Winemiller, Lopez-Fernandez, & Galetti, 2007; García-Berthou, 2001; Nurminen, Horppila, Lappalainen, & Malinen, 2003), which often retain their ability to germinate after passing through fishes' guts (Pollux, 2011). Given that fish are often highly mobile within river catchments (Lucas & Batley, 1996; Makrakis et al., 2007), this offers a potentially important vector for macrophyte dispersal. Some seeds can survive up to 36 hr in the fish gut (Horn, 1997), during which time they could be dispersed over long distances. Evidence of endozoochorus dispersal of seeds by fish has been found in Europe (Pollux, 2007; Pollux et al., 2005), North America (Chick, Cosgriff, & Gittinger, 2003; VonBank, DeBoer, Casper, & Hagy, 2018), and South America (Anderson et al., 2011: Anderson, Rojas, & Flecker, 2009), suggesting it is a widespread mechanism of upstream dispersal for river macrophytes (see Horn et al., 2011). For example, seeds of S. emersum have been found to disperse up to 27 km in the gut of the common carp (Pollux, Ouborg, Groenendael, & Klaassen, 2007), and single dispersal events by fruit-eating fish in the Amazon have been observed to transport seeds over distances greater than 5 km (Anderson et al., 2011). Fishes differ in their diets (Gerking, 1994) and propensity to move (Lucas & Baras, 2001) so fish-mediated dispersal is likely to be species-specific.

Endozoochory also occurs through water birds (Brochet et al., 2010; Charalambidou & Santamaría, 2002; Smits, Ruremonde, & Velde, 1989). Waterfowl can consume large amounts of seeds that can survive in their guts for periods of hours to days (Figuerola & Green, 2002). A recent study in Brazil showed that whole plants of the Wolffia family could survive gut passage intact (Silva et al., 2018). Given that ducks and waders can travel upwards of 50 km/ hr (Welham, 1994), there is considerable potential for long-distance dispersal (Clausen, Nolet, Fox, & Klaassen, 2002; Van Leeuwen, Velde, Groenendael, & Klaassen, 2012). There is also evidence that piscivorous birds such as cormorants can act as secondary dispersers of plant seeds (Van Leeuwen, Lovas-Kiss, Ovegård, & Green, 2017). Importantly, dispersal by birds is not restricted to river corridors, so inter-catchment transport is possible, and endozoochorus dispersal of seeds in waterfowl is possible over distances up to 3,600 km (Pollux, 2007). Bird-mediated dispersal is also thought to be responsible for gene flow between lake populations of macrophytes hundreds of kilometres apart within the Yangtze River catchment in China (Chen et al., 2009).

A number of studies have suggested the ectozoochory is uncommon in waterfowl because macrophytes propagules generally lack adherent properties, and are therefore likely to be carried only short distances (Brochet et al., 2010; Figuerola & Green, 2002; Reynolds & Cumming, 2016). However, recent studies indicate that frequent short-distance dispersal of macrophytes attached to the bodies of birds may be important. *Stepping-stone* dispersal is possible, whereby plant fragments adhere externally to birds, and are dispersed over

short distances as the birds move (Coughlan, Kelly, Davenport, & Jansen, 2017; Coughlan, Kelly, & Jansen, 2017). Although the plant fragments often tend to be moved only short distances, high frequencies of such events provide the mechanism for long-distance dispersal. Ectozoochory is likely to be particularly relevant for small macrophytes such as members of Lemnoideae (Duckweeds; Coughlan, Kelly, & Jansen, 2015; Landolt, 1986). Although not yet experimentally evaluated, it is likely that multiple short dispersal events could also result in eventual long-distance endozoochorus dispersal by fish. Irrespective of the precise mechanisms involved, it is widely accepted that zoochory is a principal mechanism for upstream dispersal of macrophytes in rivers (Coughlan, Kelly, Davenport, et al., 2017; Figuerola & Green, 2002; Pollux et al., 2006).

#### 3.3 Anemochory

Dispersal by wind (anemochory) offers an additional mechanism for propagule dispersal. Some authors have suggested this mode of dispersal is rare for aquatic macrophytes as their seeds tend to be relatively heavy, and they generally lack adaptations to promote wind dispersal (Barrat-Segretain, 1996). For instance, 90% of sedge grass (Carex sp.) seeds were deposited within 2 m of the source plant, perhaps suggesting anemochory is of limited importance in long-distance dispersal (Soomers et al., 2013). However, Soons (2006) showed that 46% of wetland plant species have adaptations to promote anemochory and argued it was of great importance in the dispersal of aquatic plants. Many emergent taxa such as Phragmites spp. and Typha spp. produce large numbers of small light seeds that are easily dispersed by wind (Shipley, Keddy, Moore, & Lemky, 1989; Soons, 2006). Although most wind-blown seeds tend to settle close to the source plant (e.g. >90% of Phragmites sp. within 30 m, Soomers et al., 2013), it is the small proportion of seeds on the tail of the dispersal curve that are important in long-distance dispersal (Nathan et al., 2008), and these seeds can be transported over distances of hundreds of kilometres (Soomers et al., 2013). Wind dispersal is likely to increase substantially during extreme weather events when wind speeds are highest and sampling is problematic (Nathan et al., 2008). Even rare long-distance dispersal events are important in facilitating gene flow between populations (Trakhtenbrot, Nathan, Perry, & Richardson, 2005). Also, as with zoochory, multiple and frequent short-distance dispersal events should theoretically result in long-distance dispersal via a stepping-stone effect (Saura, Bodin, & Fortin, 2014), and generate sufficient gene flow to prevent genetic differentiation within metapopulations. At a minimum, anemochory is likely to be an important primary mechanism of dispersal in many plants, whereby dispersal into flowing water creates secondary dispersal opportunities via hydrochory.

#### 3.4 **Anthropochory**

Human movements are increasingly spreading plants outside the confines of natural dispersal mechanisms (Hodkinson & Thompson, 1997; Wichmann et al., 2008). This not only allows for long-distance longitudinal movement of propagules and genes along rivers (Tero et al., 2003), but also dispersal across catchment boundaries, resulting in dispersal on a global scale (Ciotir & Freeland, 2016). Anthropochory is of particular relevance for the spread of invasive species. For instance, over 400 non-native macrophyte species are traded in Europe, most of which have the potential to become invasive (Hussner, 2008), and the ornamental plant trade is a major pathway for the spread of invasive macrophytes, both via deliberate and accidental introductions (Hussner, 2012). River users can spread plant propagules through recreational activities. For instance, macrophytes can frequently become entangled on recreational boating equipment, which can then be transported to other waterbodies (Johnson, Ricciardi, & Carlton, 2001; Kelly, Wantola, Weisz, & Yan, 2013; Rothlisberger, Chadderton, McNulty, & Lodge, 2010). In the UK. 64% of anglers and 78% of canoeists use their equipment in more than one catchment within a fortnight, most without any biosecurity measures (Anderson, White, Stebbing, Stentiford, & Dunn, 2014). Human-mediated dispersal can also result in gene flow between populations of native macrophytes that would otherwise be genetically isolated (Ciotir & Freeland, 2016).

# BARRIER IMPACTS ON MACROPHYTES

Artificial barriers alter the hydrology (Merritt & Wohl, 2002), temperature (Olden & Naiman, 2010), water chemistry (Byren & Davies, 1989), and sediment dynamics (Williams & Wolman, 1984) of running waters, often creating habitat discontinuities (Ward & Stanford, 1983) that can have profound influences on community assemblages (Parasiewicz, Schmutz, & Moog, 1998). These abiotic and biotic factors can potentially affect aquatic macrophytes in a variety of ways (Table 1 and Figure 2). Physical barriers have considerable potential to impact hydrochory and zoochory, and are therefore discussed in separate subsections below. Wind dispersal is unlikely to be significantly affected by the presence of barriers, except perhaps by the largest of dams. We found no papers that covered the effect of barriers on anemochory so this issue is not covered here. There is considerable evidence that barriers influence the distribution of invasive species, so we covered this topic separately (Section 4.3) along with human-mediated dispersal.

#### Barrier effects on hydrochory 4.1

Water velocity is a key parameter determining how far propagules disperse, and hence can have important consequences for plant community composition along rivers (Merritt & Wohl, 2006). Slow flows in impounded reaches can act as barriers for macrophyte dispersal, trapping drifting propagules, and resulting in high mortality (Jansson, Nilsson, Dynesius, & Andersson, 2000; Jansson, Nilsson, & Renöfält, 2000; Nilsson et al., 2010; Nilsson & Jansson, 1995). Reservoirs can reduce the density of drifting propagules in downstream reaches by

 TABLE 1
 Summary of barrier impacts on macrophyte dispersal and population structure

Attribute	Effect	Mechanism	References
Dispersal	Reservoirs trap hydrochorous propagules	Slow flow in impounded reach	Nilsson and Jansson (1995) Andersson et al. (2000) Jansson, Nilsson, and Renöfält (2000) Merritt and Wohl (2006) Nilsson et al. (2010)
	Reduced abundance and richness of hydrochorous seeds downstream	Slow flow in impounded reach	Andersson et al. (2000) Jansson, Nilsson, Dynesius, et al. (2000) Merritt and Wohl (2006)
		Reduced frequency and intensity of floods	Jansson, Nilsson, Dynesius, et al. (2000 Jansson, Nilsson, and Renöfält (2000)
	No effect of dams on abundance or diversity of drifting propagules downstream	Local (within-impoundment) sources contributed similar abundance and diversity of hydrochorous propagules to upstream	Jansson et al. (2005)
Population structure	Lower species richness downstream	Reduced hydrochory	Nilsson, Ekblad, et al. (1991) and Nilsson Gardfjell, et al. (1991) Merritt et al. (2010)
	Higher species richness downstream	Increased nutrients Intermediate disturbance caused by moderate hydropeaking	Benítez-Mora and Camargo (2014) Bernez et al. (2002) Bernez et al. (2004)
	No effect of barriers on species richness	Local seed sources downstream from dams contributed similar diversity to upstream	Merritt and Wohl (2006)
		Species richness was highly variable between sites limiting ability to identify weir effects	Mueller et al. (2011)
	Discontinuities in community composition	Reduced hydrochory	Merritt and Wohl (2006) Andersson et al. (2000) Jansson, Nilsson, Dynesius, et al. (2000
	Increased biomass in impounded area	Creation of lentic habitat	Ceschin et al. (2015) Tombolini et al. (2014)
		Reduced flow velocity	Vukov et al. (2018)
		Increased sedimentation	Rorslett and Johansen (1996) Vukov et al. (2018)
	Increased biomass downstream	Stable flows	Goes (2002) Ibáñez et al. (2012) Abati et al. (2016)
		Increased nutrients	Tena et al. (2017) Benítez-Mora and Camargo (2014)
	Decreased biomass downstream	Higher turbidity	García de Jalon et al. (1994)
		Stable flows	Casado et al. (1989)
Invasiveness	Increased spread of invasive species	Modified flow regimes	Greet, Cousens, & Webb, 2013 Van Geest et al. (2005)
		Increased anthropochory	Coetzee et al. (2009)
	Reduced spread of invasive species	Interrupted hydrochory	Thomason et al. (2018) Rood et al. (2010)

Note: Dams are defined as large barriers (generally > 5 m) that create large impoundments (reservoirs) upstream. Weirs are defined as smaller (<5 m) barriers with overtopping flow.

as much as 95%, and this effect can extend for several kilometres downstream of large dams (Merritt & Wohl, 2006). Compared to free-flowing rivers, rivers fragmented by large dams tend to show lower richness of drifting propagules, and dispersal rates are also often reduced (Andersson, Nilsson, & Johansson, 2000; Jansson, Nilsson, Dynesius, et al., 2000; Merritt & Wohl, 2006), although this is not always the case. For example, Jansson, Zinko, Merritt,

and Nilsson (2005) did not find any evidence to suggest that dams decreased the abundance or diversity of drifting propagules in a comparison of fragmented and free-flowing rivers. However, in this instance, the drifting propagule bank in the fragmented river was derived from local (within-impoundment) sources only (Jansson et al., 2005), suggesting that long-distance dispersal via hydrochory was compromised.

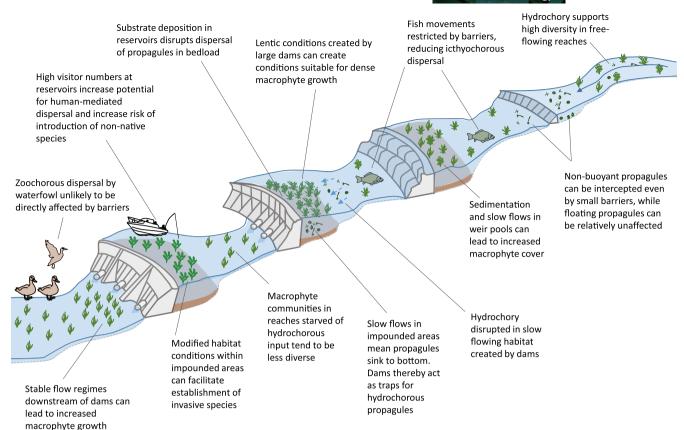


FIGURE 2 Potential impacts of barriers on macrophyte dispersal and population structure

Floods are important events for hydrochory in free-flowing rivers (Cellot, Mouillot, & Henry, 1998; Franklin et al., 2008; Gurnell, Thompson, Goodson, & Moggridge, 2008), but their intensity and frequency is reduced in many dammed rivers (Magilligan & Nislow, 2005), and this can limit hydrochorous dispersal of propagules (Jansson, Nilsson, Dynesius, et al., 2000). There is evidence that species with different dispersal strategies are affected to different degrees by flow regulation (Jansson, Nilsson, Dynesius, et al., 2000; Jansson, Nilsson, & Renöfält, 2000). For example, the reduced frequency of floods in regulated reaches can prevent transport of non-buoyant propagules, whereas those with floating propagules can show higher probability of dispersal (Jansson, Nilsson, Dynesius, et al., 2000; Jansson, Nilsson, & Renöfält, 2000).

Barrier design can influence the extent to which hydrochory is disrupted. Large dams with big reservoirs are likely to have a greater impact on hydrochory than smaller barriers with negligible impoundments. Throughflow barriers (e.g. culverts) and overflow (e.g. weirs) barriers are likely to intercept less propagules than bottom-release dams. Impoundments with thick vegetative growth should intercept more drifting propagules than sparely vegetated impoundments (Riis, 2008). Structures with sediment release mechanisms should cause less disruption to transport of seeds in the bedload (Markwith & Leigh, 2008). Hence, the impact of barriers on hydrochory is highly context-dependent (Figure 3).

The impact of low-head barriers on macrophytes has been largely unexplored compared to the effects of large dams. Although their

impact is likely to be less severe than large dams, smaller barriers such as weirs modify river flows, often creating slow velocity areas (weir pools) immediately upstream. These weir pools tend to stabilise the substrate and increase settlement of fine sediments (Merritt & Wohl, 2006). In trapping river substrates, small barriers probably prevent or at least significantly reduce movement of non-buoyant seeds in the bedload (Markwith & Leigh, 2008, 2012). Stable substrates create opportunities for the establishment of macrophytes that would otherwise have been unable to root (Riis & Biggs, 2003) and can further exacerbate changes by creating a positive feedback loop, whereby the presence of standing macrophytes increases sedimentation rates (Gurnell et al., 2006; Jones, Collins, Naden, & Sear, 2012; Sand-Jensen et al., 1989) and thus increases propagule settlement (Gurnell et al., 2008; Riis, 2008). For example, in Norway, weirs are commonly built as part of small-scale hydropower schemes, but weir pools are often associated with increased siltation and subsequent growth of macrophytes, which are regarded as a nuisance (Rorslett & Johansen, 1996). However, other studies have found little evidence that weirs affected macrophyte diversity or abundance (Mueller et al., 2011). The variation in the response of macrophytes is likely to depend on the nature of hydrological alteration: i.e. where weirs stabilise flows and substrate macrophyte cover tends to increase, whereas barriers with negligible effects on hydrology and substrate movement tend to have little effect on macrophytes.

The potential link between disrupted hydrochory and community structure downstream is unclear. Although dams can significantly

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### Characteristic **Factor** Medium Large Absent Small **Barrier height** Flow through (e.g. culvert) Overflow (e.g. weir) Bottom release (e.g. dam) Barrier design Negligible impoundment Small (e.g. weir pool) Large reservoir Impoundment size Slow, laminar Fast, turbulent Flow within impoundment Severely disrupted Uninterrupted **Bedload movement** Sparse Dense Vegetation within impoundment Low flow High flow River state Sinking propagules Propagule type Buoyant propagules **Hydrochory** Negligible impact on hydrochory severely disrupted

FIGURE 3 The influence of various context-dependent factors on the impact of barriers on hydrochory

reduce hydrochorous dispersal, the abundance and diversity of plant populations downstream of dams may in some cases remain the same as upstream (Merritt & Wohl, 2006). Discontinuities in community composition have been associated with dams, with assemblages exhibiting a shift from a composition similar to the drifting hydrochorous propagule bank upstream, to communities derived from local seed-bearing plants downstream (Andersson et al., 2000; Jansson, Nilsson, Dynesius, et al., 2000). Decreases in macrophyte diversity and abundance have been reported downstream of dams (Casado, García de Jalon, Delolmo, Barcelo, & Menes, 1989; García de Jalon, Sanchez, & Camargo, 1994), while in other cases an increase in macrophyte abundance has been reported (Abati, Minciardi, Ciadamidaro, Fattorini, & Ceschin, 2016; Goes, 2002; Tena, Vericat, Gonzalo, & Batalla, 2017), although in these studies, changes in macrophyte population structure were not directly linked to disruptions of hydrochory, and probably related to differences in hydrological regime. However, other studies have found evidence that the richness of riverine plant communities was linked to hydrochorous seed input, with free-flowing rivers showing higher richness (Merritt, Nilsson, & Jansson, 2010; Nilsson, Ekblad, et al., 1991; Nilsson, Gardfjell, et al., 1991).

Many of the observed changes in macrophyte community have been associated with the hydrological effects of dams, rather than their role in disrupting hydrochory. Stable flow conditions often found downstream of dams can increase aquatic plant cover in affected reaches (Abati et al., 2016; Goes, 2002; Ibáñez, Caiola, Rovira, & Real, 2012). Moderate disturbance caused by hydropeaking (frequent, short duration, artificial flow events) can also lead to increased macrophyte richness and abundance, compared to unregulated rivers (Bernez, Daniel, Haury, & Ferreira, 2004; Bernez, Haury, & Ferreira, 2002). Where hydrological disturbance is more severe, macrophyte

communities tend to show low diversity and be less abundant (Casado et al., 1989; García de Jalon et al., 1994; Merritt et al., 2010).

Impoundments upstream of dams are characterised by slow flows, reduced turbulence and more uniform habitats, increasing sedimentation rates and creating conditions that resemble lentic systems (Anderson, Moggridge, Warren, & Shucksmith, 2015; Vukov et al., 2018). Dissolved concentrations of critical nutrients such as phosphorous and nitrate are often higher in these impounded reaches, leading to increased plant growth (Benítez-Mora & Camargo, 2014). As a result of these changes, slow-flowing habitats immediately upstream of dams often support high macrophyte biomass, albeit generally with communities more representative of lacustrine habitats (Ceschin et al., 2015; Tombolini, Caneva, Cancellieri, Abati, & Ceschin, 2014; Vukov et al., 2018). Some authors have described these changes as favourable due to locally increased macrophyte diversity (Ceschin et al., 2015). However, the establishment of largely lacustrine macrophyte species probably occurs to the detriment of riverine flora and fauna within impounded areas.

# 4.2 | Barrier effects on zoochory

Given the importance of icthyochory (movement of seeds by fish) for upstream dispersal (Anderson et al., 2011; Horn et al., 2011; Pollux et al., 2006), impediment of fish movements by barriers (Garcia de Leaniz, 2008; Lucas & Batley, 1996; Winter & Van Densen, 2001) could potentially impact macrophyte dispersal and population connectivity. To date, no study has specifically assessed how the presence of barriers may affect endozoochorous dispersal of seeds by fish, although it has been raised as an issue of concern (Correa et al.,

2007; Horn et al., 2011). The group of fish in which seed dispersal has been identified tend to be weaker-swimming members of river fish communities such as cyprinids, characids, and ictalurids (Anderson et al., 2009: Chick et al., 2003: VonBank, DeBoer, et al., 2018). These species are more likely to be affected by barriers because they lack the swimming speed and leaping ability to overcome many obstacles (Beecham, 2004; Langerhans & Reznick, 2010; Tudorache, Viaene, Blust, Vereecken, & Boeck, 2008).

The presence of river barriers is unlikely to affect the movements of birds directly, but changes in the distribution of riverine habitats brought about by flow regulation can alter the composition and distribution of waterfowl communities (Nilsson & Dynesius, 1994). For example, the accumulation of fish at barriers can also lead to local increases in piscivorous birds (Baumgartner, Stuart, & Zampatti, 2008; Stevens et al., 1997) and still waters within impoundments provide habitat for many waterfowl (Nilsson & Dynesius, 1994). Because birds can act as important agents of propagule dispersal (Charalambidou & Santamaría, 2002; Coughlan et al., 2015; Figuerola & Green, 2002), changes in their distribution have the potential to influence aquatic plant dispersal, although this has not yet been examined. Further research is required to elucidate the impacts of barriers for zoochorous dispersal of seeds by animals.

#### 4.3 Barrier effects on invasive macrophytes

Hydrochory has been highlighted as an important mechanism for the spread of aquatic invasive plants (Aronson, Patel, O'Neill, & Ehrenfeld, 2017; Okada, Grewell, & Jasieniuk, 2009; Thébaud & Debussche, 1991). The trapping of drifting propagules by large barriers such as dams can inhibit or prevent the spread of invasive species that rely on hydrochory for dispersal (Rood, Braatne, & Goater, 2010). However, any such effect is also likely to impact population connectivity of native macrophytes (Merritt & Wohl, 2006; Nilsson et al., 2010).

Invasive species tend to be most successful where naturally occurring communities are stressed by anthropogenic disturbance (Byers et al., 2002; Johnson, Olden, & Vander Zanden, 2008; Strayer, 2010), including damming (Greet et al., 2013; Johnson et al., 2008). Hydrological modifications associated with river barriers can result in changes in community composition, as native macrophytes may be unable to cope with modified conditions (Catford & Jansson, 2014), creating opportunities for invasive macrophytes to establish. For example, damming of the river Guadiana in Spain has led to increased spread of the invasive water hyacinth (Eichhornia crassipes; Téllez et al., 2008). A recent study has also shown that thick mats of water hyacinth can themselves trap and disrupt downstream transport of hydrochorous seeds (Vonbank, Casper, Pendleton, & Hagy, 2018). Artificial reservoirs in North America tend to support more invasive species than natural lakes, probably because native species have less of a stronghold in artificial systems (Johnson et al., 2008). Flow regulation in the River Rhine has favoured the spread of invasive Elodea nuttallii, which has become dominant in many altered reaches (Van Geest, Coops, Roijackers, Buijse, & Scheffer, 2005) and modified flow regimes downstream of dams have also favoured the invasion of non-native macrophytes in riverine wetlands within the Murray River, Australia (Catford, Downes, Gippel, & Vesk, 2011). However, a reduction in flood disturbance due to river regulation in a Californian river system resulted in reduced propagule dispersal in the invasive aquatic macrophyte Ludwigia hexapetala (Thomason, McCort, Netherland, & Grewell, 2018). These contrasting findings indicate that the outcomes of hydrological modification depend on the flow regimes imposed and the dispersal traits of the invasive nlants present

Reservoirs often receive higher numbers of boat users and anglers than free-flowing sections of rivers (Cooper, 2006; Havel, Lee, & Vander Zanden, 2005), and impounded areas can be sites of high introduction risk within catchments (Jacobs & Macisaac, 2009; Johnson et al., 2008: Tamavo & Olden, 2014). For instance, artificial reservoirs in South Africa were highlighted as high-risk areas for invasion of Hydrilla venticillata due to high boat traffic (Coetzee, Hill, & Schlange, 2009). Recreational disturbance (e.g. kayaking, boaters, and fishermen) in impounded reaches can also result in the fragmentation of invasive plants, resulting in higher hydrochorous dispersal (Thomason et al., 2018). Outreach efforts to increase public awareness, biosecurity campaigns, and promotion of rigorous cleaning protocols can be highly effective in reducing anthropogenic dispersal of invasive macrophytes (Rothlisberger et al., 2010).

# CONCLUSIONS

Macrophytes are an essential component of healthy rivers and barriers have the potential for impacting them in subtle, insidious ways (Pringle, 2001, 2003), and yet, the effects of anthropogenic barriers on river macrophytes have received little attention compared to fish and other riverine biota. For instance, while we found 333 articles in Web of Science dealing with barrier impacts on fish, and 30 on macroinvertebrates, only 19 investigated effects on macrophytes (see Table S3 for search strings). This is perhaps due to the assumption that macrophyte populations are sedentary, and therefore relatively unaffected by barriers. However, there is strong evidence that macrophytes can disperse over relatively long distances, both drifting with the river flow and transported via animal movement, and this dispersal is crucial for maintaining population connectivity and persistence.

The empirical evidence indicates large barriers such as dams have substantial impacts on macrophyte dispersal (Jansson, Nilsson, & Renöfält, 2000; Nilsson et al., 2010), acting as traps to drifting propagules, and thereby starving downstream reaches of hydrochorous input (Andersson et al., 2000; Merritt & Wohl, 2006). However, the abundance and diversity of the drifting propagule bank are not always affected downstream of dams (Jansson et al., 2005), and the consequences of disrupted hydrochory for community dynamics are unclear. The existing literature has overwhelmingly focussed on large dams only, despite the fact that small weirs and low-head

TABLE 2 Knowledge gaps identified during this review

Suggested future research directions	Relevant studies
The dominant role of hydrochory in downstream dispersal is well established, but further research is required to identify mechanisms supporting upstream dispersal, and their relative importance	Charalambidou and Santamaría (2002), Markwith and Scanlon (2007), Pollux et al. (2009), and Pollux et al. (2005),
There is contradictory evidence regarding the consequences of interrupted hydrochory for the composition of macrophyte communities within rivers. Little is known about the site-specific factors influencing barrier effects on macrophyte population dynamics	Andersson et al. (2000), Jansson et al. (2005), Merritt and Wohl (2002, 2006), and Nilsson et al. (2010)
Many animal species can disperse macrophytes, particularly birds and fish, but the prevalence of animal dispersal is uncertain	Charalambidou and Santamaría (2002), Figuerola and Green (2002), Pollux et al. (2006), and Pollux et al. (2005)
Given the well documented effects that river barriers have on fish movements, their impact on icthyochory requires evaluation	Correa et al. (2007), Horn et al. (2011), Pollux et al. (2006), and Pollux et al. (2005)
Stepping-stone dispersal is likely to be of great importance in supporting long-distance dispersal by both zoochory and anemochory, and these processes require further investigation	Coughlan, Kelly, Davenport, et al. (2017), Coughlan et al. (2015), Coughlan, Kelly, and Jansen (2017), and Saura et al. (2014)
There is good evidence that the presence of barriers can result in the establishment of invasive macrophytes, but the mechanisms facilitating colonisation are not well understood and need evaluation	Catford et al. (2011), Johnson et al. (2008), and Rood et al. (2010)
Most studies examining barrier effects on macrophytes have focussed on large dams and reservoirs, but low-head barriers are much more numerous, and their cumulative impact could be considerable and requires assessment	Mueller et al. (2011), Markwith and Leigh (2008), and Rorslett and Johansen (1996)
Mitigation solutions for reducing the impact of river barriers on macrophyte populations are unavailable and require investigation	N/A

structures are much more abundant (Garcia de Leaniz et al., 2018; Januchowski-Hartley et al., 2013; Jones et al., 2019), and may also impact on macrophytes (Rorslett & Johansen, 1996). The potential for small barriers to intercept hydrochloric drift has not been adequately explored (Table 2).

Most studies assessing barrier effects on macrophyte dispersal have focussed on their influence on hydrochory. However, upstream dispersal is equally critical to maintaining macrophyte abundance and distribution, and the role of barriers in disrupting potentially important mechanisms such as icthyochory needs to be examined (Table 2). This will probably require knowledge of the spatial scale of zoochory, the precise dispersal mechanisms involved (e.g. stepping-stone effects), and the extent to which barriers effect the movements and distributions of the specific taxa that act as dispersal vectors (Table 2).

There is contradictory evidence regarding damming effects on the macrophyte standing crop in affected reaches, with some studies reporting a negative impact on macrophyte populations (Casado et al., 1989; Nilsson, Ekblad, et al., 1991; Nilsson, Gardfjell, et al., 1991), while others indicate increases in abundance and diversity in regulated reaches (Ceschin et al., 2015; Vukov et al., 2018). Hence, the effect of dams on macrophytes is complex and appears to be very much context-dependent. The local factors influencing the impact of barriers on macrophytes require further investigation (Table 2). Some studies indicate small barriers such as weirs can cause shifts in macrophyte distribution and abundance (Rorslett & Johansen, 1996) while others report non-significant effects (Mueller et al., 2011), and these contrasting outcomes may depend on the

nature of hydrological alterations. However, the impact of small barriers on macrophyte population dynamics requires further attention (Table 2). Also, flow regulation appears to favour the establishment of invasive macrophytes, but the mechanisms involved are not well understood, and need evaluation (Table 2).

Even common plant species can be susceptible to genetic impoverishment due to habitat fragmentation (Honnay & Jacquemyn, 2007). River fragmentation is an ongoing process (Couto & Olden, 2018; Grill et al., 2015), so increasing isolation of populations could potentially leave many macrophyte species vulnerable to genetic erosion. In terrestrial plants, low levels of gene flow (1 seed per generation) are sufficient to prevent genetic differentiation between populations (Honnay, Jacquemyn, Bossuyt, & Hermy, 2005; Wright, 1931). However, there is a need to identify the frequency of dispersal events over dams that would be required in order to avoid genetic divergence and population decline in river macrophytes (Table 2). This would probably require metabarcoding to examine rates of gene flow under different frequencies of hydrochloric immigration, including multi-generational studies to assess rates of genetic divergence under different dispersal scenarios. Such studies would need to be undertaken both at catchment and sub-catchment scales. There is also little knowledge of what modifications might be made to dam and reservoirs in order to improve dispersal of macrophyte propagules (Table 2). Due to the different dispersal characteristics of distinct taxa (e.g. buoyant and sinking propagules), a variety of alterations would probably be required to provide for uninterrupted dispersal of diverse macrophyte communities.

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# **REFERENCES**

- Abati, S., Minciardi, M. R., Ciadamidaro, S., Fattorini, S., & Ceschin, S. (2016). Response of macrophyte communities to flow regulation in mountain streams. Environmental Monitoring and Assessment, 188(7), 414. https://doi.org/10.1007/s10661-016-5420-2
- Anderson, D., Moggridge, H., Warren, P., & Shucksmith, J. (2015). The impacts of 'run-of-river' hydropower on the physical and ecological condition of rivers. Water and Environment Journal, 29(2), 268-276. https://doi.org/10.1111/wej.12101
- Anderson, J. T., Nuttle, T., Saldaña Rojas, J. S., Pendergast, T. H., & Flecker, A. S. (2011). Extremely long-distance seed dispersal by an overfished Amazonian frugivore. Proceedings of the Royal Society B, 278, 3329-3335. https://doi.org/10.1098/rspb.2011.0155
- Anderson, J. T., Rojas, J. S., & Flecker, A. S. (2009). High-quality seed dispersal by fruit-eating fishes in Amazonian floodplain habitats. Oecologia, 161(2), 279-290. https://doi.org/10.1007/ s00442-009-1371-4
- Anderson, L. G., White, P. C., Stebbing, P. D., Stentiford, G. D., & Dunn, A. M. (2014). Biosecurity and vector behaviour: Evaluating the potential threat posed by anglers and canoeists as pathways for the spread of invasive non-native species and pathogens. PLoS ONE, 9(4), e92788. https://doi.org/10.1371/journal.pone.0092788
- Andersson, E., Nilsson, C., & Johansson, M. E. (2000). Effects of river fragmentation on plant dispersal and riparian flora. Regulated Rivers: Research & Management, 16(1), 83-89. https://doi.org/10.1002/ (SICI)1099-1646(200001/02)16:1<83:AID-RRR567>3.0.CO;2-T
- Aronson, M. F., Patel, M. V., O'Neill, K. M., & Ehrenfeld, J. G. (2017). Urban riparian systems function as corridors for both native and invasive plant species. Biological Invasions, 19(12), 3645-3657. https:// doi.org/10.1007/s10530-017-1583-1
- Barrat-Segretain, M. H. (1996). Strategies of reproduction, dispersion, and competition in river plants: A review. Vegetatio, 123(1), 13-37. https://doi.org/10.1007/BF00044885
- Baumgartner, L. J., Stuart, I. G., & Zampatti, B. P. (2008). Determining diel variation in fish assemblages downstream of three weirs in a regulated lowland river. Journal of Fish Biology, 72(1), 218-232. https:// doi.org/10.1111/j.1095-8649.2007.01696.x
- Beecham, R. V. (2004). A study of the swimming capabilities of blue, Ictalurus furcatus, and channel, I. punctatus, catfish. The University of Mississippi.
- Benítez-Mora, A., & Camargo, J. A. (2014). Ecological responses of aquatic macrophytes and benthic macroinvertebrates to dams in the Henares River Basin (Central Spain). Hydrobiologia, 728(1), 167-178. https://doi.org/10.1007/s10750-014-1816-6

- Bernez, I., Daniel, H., Haury, J., & Ferreira, M. T. (2004). Combined effects of environmental factors and regulation on macrophyte vegetation along three rivers in Western France. River Research and Applications, 20(1), 43-59. https://doi.org/10.1002/rra.718
- Bernez, I., Haury, J., & Ferreira, M. T. (2002). Downstream effects of a hydroelectric reservoir on aquatic plant assemblages. The Scientific World Journal, 2, 740-750. https://doi.org/10.1100/tsw.2002.142
- Biggs, B. J. (1996). Hydraulic habitat of plants in streams. River Research and Applications, 12(2-3), 131-144. https://doi.org/10.1002/ (SICI)1099-1646(199603)12:2/3<131:AID-RRR385>3.0.CO;2-X
- Boedeltje, G., Bakker, J. P., Brinke, A. T., Van Groenendael, J. M., & Soesbergen, M. (2004). Dispersal phenology of hydrochorous plants in relation to discharge, seed release time and buoyancy of seeds: The flood pulse concept supported. Journal of Ecology, 92(5), 786-796. https://doi.org/10.1111/j.0022-0477.2004.00906.x
- Branco, P., Amaral, S. D., Ferreira, M. T., & Santos, J. M. (2017). Do small barriers affect the movement of freshwater fish by increasing residency? Science of the Total Environment, 581, 486-494. https://doi. org/10.1016/j.scitotenv.2016.12.156
- Brochet, A. L., Guillemain, M., Fritz, H., Gauthier-Clerc, M., & Green, A. J. (2010). Plant dispersal by teal (Anas crecca) in the Camargue: Duck guts are more important than their feet. Freshwater Biology, 55(6), 1262-1273. https://doi.org/10.1111/j.1365-2427.2009.02350.x
- Bunn, S. E., Davies, P. M., Kellaway, D. M., & Prosser, I. P. (1998). Influence of invasive macrophytes on channel morphology and hydrology in an open tropical lowland stream, and potential control by riparian shading. Freshwater Biology, 39(1), 171-178. https://doi. org/10.1046/j.1365-2427.1998.00264.x
- Byers, J. E., Reichard, S., Randall, J. M., Parker, I. M., Smith, C. S., Lonsdale, W. M., ... Hayes, D. (2002). Directing research to reduce the impacts of nonindigenous species. Conservation Biology, 16(3), 630-640. https://doi.org/10.1046/j.1523-1739.2002.01057.x
- Byren, B. A., & Davies, B. R. (1989). The effect of stream regulation on the physicochemical properties of the Palmiet River, South Africa. Regulated Rivers: Research and Management, 3, 107–121.
- Carpenter, S. R., & Lodge, D. M. (1986). Effects of submersed macrophytes on ecosystem processes. Aquatic Botany, 26, 341-370. https://doi.org/10.1016/0304-3770(86)90031-8
- Casado, C., García de Jalon, D., Delolmo, C. M., Barcelo, E., & Menes, F. (1989). The effect of an irrigation and hydroelectric reservoir on its downstream communities. Regulated Rivers: Research & Management, 4(3), 275-284. https://doi.org/10.1002/rrr.3450040306
- Catford, J. A., Downes, B. J., Gippel, C. J., & Vesk, P. A. (2011). Flow regulation reduces native plant cover and facilitates exotic invasion in riparian wetlands. Journal of Applied Ecology, 48(2), 432-442. https:// doi.org/10.1111/j.1365-2664.2010.01945.x
- Catford, J. A., & Jansson, R. (2014). Drowned, buried and carried away: Effects of plant traits on the distribution of native and alien species in riparian ecosystems. New Phytologist, 204(1), 19-36. https://doi. org/10.1111/nph.12951
- Cellot, B., Mouillot, F., & Henry, C. P. (1998). Flood drift and propagule bank of aquatic macrophytes in a riverine wetland. Journal of Vegetation Science, 9(5), 631-640. https://doi.org/10.2307/3237281
- Ceschin, S., Tombolini, I., Abati, S., & Zuccarello, V. (2015). The effect of river damming on vegetation: Is it always unfavourable? A case study from the River Tiber (Italy). Environmental Monitoring and Assessment, 187(5), 301. https://doi.org/10.1007/s10661-015-4521-7
- Chambers, P. A., Lacoul, P., Murphy, K. J., & Thomaz, S. M. (2008). Global diversity of aquatic macrophytes in freshwater. Hydrobiologia, 595(1), 9-26. https://doi.org/10.1007/s10750-007-9154-6
- Charalambidou, I., & Santamaría, L. (2002). Waterbirds as endozoochorous dispersers of aquatic organisms: A review of experimental evidence. Acta Oecologica, 23(3), 165-176. https://doi.org/10.1016/ S1146-609X(02)01148-7

- Chen, Y., Li, X., Yin, L., Cheng, Y., & Li, W. (2009). Genetic diversity and migration patterns of the aquatic macrophyte Potamogeton malaianus in a potamo-lacustrine system. Freshwater Biology, 54(6), 1178–1188.
- Chick, J. H., Cosgriff, R. J., & Gittinger, L. S. (2003). Fish as potential dispersal agents for floodplain plants: First evidence in North America. Canadian Journal of Fisheries and Aquatic Sciences, 60(12), 1437–1439. https://doi.org/10.1139/f03-155
- Ciotir, C., & Freeland, J. (2016). Cryptic intercontinental dispersal, commercial retailers, and the genetic diversity of native and non-native cattails (*Typha* spp.) in North America. *Hydrobiologia*, 768(1), 137–150. https://doi.org/10.1007/s10750-015-2538-0
- Clarke, S. J., & Wharton, G. (2001). Sediment nutrient characteristics and aquatic macrophytes in lowland English rivers. Science of the Total Environment, 266(1), 103–112. https://doi.org/10.1016/S0048-9697(00)00754-3
- Clausen, P., Nolet, B. A., Fox, A. D., & Klaassen, M. (2002). Long-distance endozoochorous dispersal of submerged macrophyte seeds by migratory waterbirds in northern Europe—A critical review of possibilities and limitations. *Acta Oecologica*, 23(3), 191–203. https://doi.org/10.1016/S1146-609X(02)01150-5
- Coetzee, J. A., Hill, M. P., & Schlange, D. (2009). Potential spread of the invasive plant *Hydrilla verticillata* in South Africa based on anthropogenic spread and climate suitability. *Biological Invasions*, 11(4), 801– 812. https://doi.org/10.1007/s10530-008-9294-2
- Cooper, C. (2006).Lakes as tourism destination resources. Lake Tourism. An integrated approach to lacustrine tourism systems, 27–42.
- Correa, S. B., Winemiller, K. O., Lopez-Fernandez, H., & Galetti, M. (2007). Evolutionary perspectives on seed consumption and dispersal by fishes. AIBS Bulletin, 57(9), 748–756. https://doi.org/10.1641/B570907
- Coughlan, N. E., Kelly, T. C., Davenport, J., & Jansen, M. A. K. (2017). Up, up and away: Bird-mediated ectozoochorous dispersal between aquatic environments. *Freshwater Biology*, 62(4), 631–648. https://doi.org/10.1111/fwb.12894
- Coughlan, N. E., Kelly, T. C., & Jansen, M. A. K. (2015). Mallard duck (Anas platyrhynchos)-mediated dispersal of Lemnaceae: A contributing factor in the spread of invasive Lemna minuta? Plant Biology, 17(Suppl. 1), 108–114.
- Coughlan, N. E., Kelly, T. C., & Jansen, M. A. K. (2017). "Step by step": High frequency short-distance epizoochorous dispersal of aquatic macrophytes. *Biological Invasions*, 19(2), 625–634. https://doi.org/10.1007/s10530-016-1293-0
- Couto, T. B., & Olden, J. D. (2018). Global proliferation of small hydropower plants-science and policy. Frontiers in Ecology and the Environment, 16(2), 91-100. https://doi.org/10.1002/fee.1746
- Dawson, F. H. (1988). Water flow and the vegetation of running water.
  In J. J. Symoens (Ed.), Vegetation of inland waters (pp. 283–309).
  Dordrecht, the Netherlands: Kluwer Academic Publishers.
- Dehais, C., Eudeline, R., Berrebi, P., & Argillier, C. (2010). Microgeographic genetic isolation in chub (Cyprinidae: *Squalius cephalus*) population of the Durance river: Estimating fragmentation by dams. *Ecology of Freshwater Fish*, 19(2), 267–278. https://doi.org/10.1111/j.1600-0633.2010.00411.x
- Diebel, M. W., Fedora, M., Cogswell, S., & O'Hanley, J. R. (2015). Effects of road crossings on habitat connectivity for stream-resident fish. *River Research and Applications*, 31(10), 1251–1261. https://doi.org/10.1002/rra.2822
- Dudgeon, D., Arthington, A. H., Gessner, M. O., Kawabata, Z.-I., Knowler,
  D. J., Lévêque, C., ... Sullivan, C. A. (2006). Freshwater biodiversity:
  Importance, threats, status and conservation challenges. *Biological Reviews*, 81(2), 163–182. https://doi.org/10.1017/S146479310 5006950
- Ellis, L. E., & Jones, N. E. (2013). Longitudinal trends in regulated rivers: A review and synthesis within the context of the

- serial discontinuity concept. Environmental Reviews, 21(3), 136–148. https://doi.org/10.1139/er-2012-0064
- European Commission (2000). Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000 establishing a framework for the community action in the field of water policy. Official Journal of the European Commission Legis, 327, 1–72.
- Fagan, W. F. (2002). Connectivity, fragmentation, and extinction risk in dendritic metapopulations. *Ecology*, 83(12), 3243–3249. https://doi. org/10.1890/0012-9658(2002)083[3243:CFAERI]2.0.CO;2
- Figuerola, J., & Green, A. J. (2002). Dispersal of aquatic organisms by waterbirds: A review of past research and priorities for future studies. *Freshwater Biology*, 47, 483–494. https://doi.org/10.1046/j.1365-2427.2002.00829.x
- Fleming, J. P., & Dibble, E. D. (2015). Ecological mechanisms of invasion success in aquatic macrophytes. *Hydrobiologia*, 746(1), 23–37. https://doi.org/10.1007/s10750-014-2026-y
- Franklin, P., Dunbar, M., & Whitehead, P. (2008). Flow controls on lowland river macrophytes: A review. *Science of the Total Environment*, 400(1–3), 369–378. https://doi.org/10.1016/j.scitotenv.2008.06.018
- Fullerton, A. H., Burnett, K. M., Steel, E. A., Flitcroft, R. L., Pess, G. R., Feist, B. E., ... Sanderson, B. L. (2010). Hydrological connectivity for riverine fish: Measurement challenges and research opportunities. Freshwater Biology, 55(11), 2215–2237. https://doi.org/10.1111/j.1365-2427.2010.02448.x
- García de Jalon, D. G., Sanchez, P., & Camargo, J. A. (1994). Downstream effects of a new hydropower impoundment on macrophyte, macroinvertebrate and fish communities. *Regulated Rivers: Research & Management*, 9(4), 253–261. https://doi.org/10.1002/rrr.34500 90406
- Garcia de Leaniz, C. (2008). Weir removal in salmonid streams: Implications, challenges and practicalities. *Hydrobiologia*, 609, 83–96. https://doi.org/10.1007/s10750-008-9397-x
- Garcia de Leaniz, C., Belletti, B., Bizzi, S., Segura, G., Börger, L., Jones, J., ....The AMBER Consortium (2018). The Importance of having a good database for restoring river connectivity: The AMBER Barrier Atlas. In K. Brink, P. Gough, J. Royte, P. P. Schollema, & H. Wanningen (Eds.), From sea to source v2 protection and restoration of fish migration in rivers worldwide (pp. 142–145). World Fish Migration Foundation.
- García-Berthou, E. (2001). Size- and depth-dependent variation in habitat and diet of the common carp (*Cyprinus carpio*). *Aquatic Sciences*, 63, 466–476.
- Gerking, S. D. (1994). Feeding ecology of fish. San Diego, CA: Academic Press.
- Goes, B. J. M. (2002). Effects of river regulation on aquatic macrophyte growth and floods in the Hadejia-Nguru Wetlands and flow in the Yobe River, northern Nigeria; implications for future water management. River Research and Applications, 18(1), 81–95. https://doi.org/10.1002/rra.633
- Gornall, R. J., Hollingsworth, P. M., & Preston, C. D. (1998). Evidence for spatial structure and directional gene flow in a population of an aquatic plant *Potamogeton coloratus*. *Heredity*, 80, 414–421. https:// doi.org/10.1046/j.1365-2540.1998.00270.x
- Greet, J., Cousens, R. D., & Webb, J. A. (2013). More exotic and fewer native plant species: Riverine vegetation patterns associated with altered seasonal flow patterns. River Research and Applications, 29(6), 686–706. https://doi.org/10.1002/rra.2571
- Grenouillet, G., Pont, D., & Seip, K. L. (2002). Abundance and species richness as a function of food resources and vegetation structure: Juvenile fish assemblages in rivers. *Ecography*, 25(6), 641–650. https://doi.org/10.1034/j.1600-0587.2002.250601.x
- Grill, G., Lehner, B., Lumsdon, A. E., MacDonald, G. K., Zarfl, C., & Liermann, C. R. (2015). An index-based framework for assessing patterns and trends in river fragmentation and flow regulation by global dams at multiple scales. *Environmental Research Letters*, 10(1), 015001. https://doi.org/10.1088/1748-9326/10/1/015001

- Gurnell, A. M., Thompson, K., Goodson, J., & Moggridge, H. (2008).
  Propagule deposition along river margins: Linking hydrology and ecology. *Journal of Ecology*, 96(3), 553–565. https://doi.org/10.1111/j.1365-2745.2008.01358.x
- Gurnell, A. M., Van Oosterhout, M. P., De Vlieger, B., & Goodson, J. M. (2006). Reach-scale interactions between aquatic plants and physical habitat: River Frome, Dorset. River Research and Applications, 22(6), 667–680. https://doi.org/10.1002/rra.929
- Havel, J. E., Lee, C. E., & Vander Zanden, J. M. (2005). Do reservoirs facilitate invasions into landscapes? *BioScience*, 55(6), 518–525. https://doi.org/10.1641/0006-3568(2005)055[0518:DRFIIL]2.0.CO;2
- Hodkinson, D. J., & Thompson, K. (1997). Plant dispersal: The role of man. Journal of Applied Ecology, 34(6), 1484–1496. https://doi. org/10.2307/2405264
- Honnay, O., & Jacquemyn, H. (2007). Susceptibility of common and rare plant species to the genetic consequences of habitat fragmentation. *Conservation Biology*, 21(3), 823–831. https://doi. org/10.1111/j.1523-1739.2006.00646.x
- Honnay, O., Jacquemyn, H., Bossuyt, B., & Hermy, M. (2005). Forest fragmentation effects on patch occupancy and population viability of herbaceous plant species. *New Phytologist*, 166(3), 723–736. https://doi.org/10.1111/j.1469-8137.2005.01352.x
- Honnay, O., Jacquemyn, H., Nackaerts, K., Breyne, P., & Van Looy, K. (2010). Patterns of population genetic diversity in riparian and aquatic plant species along rivers. *Journal of Biogeography*, *37*(9), 1730–1739. https://doi.org/10.1111/j.1365-2699.2010.02331.x
- Horn, M. H. (1997). Evidence for dispersal of fig seeds by the fruit-eating characid fish *Brycon guatemalensis* Regan in a Costa Rican tropical rain forest. *Oecologia*, 109, 259–264. https://doi.org/10.1007/s004420050081
- Horn, M. H., Correa, S. B., Parolin, P., Pollux, B., Anderson, J. T., Lucas, C., ... Goulding, M. (2011). Seed dispersal by fishes in tropical and temperate fresh waters: The growing evidence. *Acta Oecologica*, 37(6), 561–577. https://doi.org/10.1016/j.actao.2011.06.004
- Horreo, J. L., Martinez, J. L., Ayllon, F., Pola, I. G., Monteoliva, J. A., Héland, M., & Garcia Vasquez, E. (2011). Impact of habitat fragmentation on the genetics of populations in dendritic landscapes. *Freshwater Biology*, 56(12), 2567–2579. https://doi.org/10.1111/j.1365-2427.2011.02682.x
- Horvath, T. G. (2004). Retention of particulate matter by macrophytes in a first-order stream. *Aquatic Botany*, 78(1), 27–36. https://doi.org/10.1016/j.aquabot.2003.09.003
- Hussner, A. (2008). Zur Okologie und Okophysiologie aquatischer Neophyten in Nordrhein-Westfalen. PhD thesis, Hein-rich-Heine-University, Dusseldorf, Germany.
- Hussner, A. (2012). Alien aquatic plant species in European countries. *Weed Research*, 52(4), 297–306. https://doi.org/10.1111/j.1365-3180.2012.00926.x
- Ibáñez, C., Caiola, N., Rovira, A., & Real, M. (2012). Monitoring the effects of floods on submerged macrophytes in a large river. Science of the Total Environment, 440, 132–139. https://doi.org/10.1016/j.scitotenv.2012.07.073
- Jacobs, M. J., & Macisaac, H. J. (2009). Modelling spread of the invasive macrophyte *Cabomba caroliniana*. Freshwater Biology, 54(2), 296–305.
- Jansson, R., Nilsson, C., Dynesius, M., & Andersson, E. (2000). Effects of river regulation on river-margin vegetation: A comparison of eight boreal rivers. *Ecological Applications*, 10(1), 203–224. https://doi. org/10.1890/1051-0761(2000)010[0203:EORROR]2.0.CO;2
- Jansson, R., Nilsson, C., & Renöfält, B. (2000). Fragmentation of riparian floras in rivers with multiple dams. *Ecology*, 81(4), 899–903. https:// doi.org/10.1890/0012-9658(2000)081[0899:FORFIR]2.0.CO;2
- Jansson, R., Zinko, U., Merritt, D. M., & Nilsson, C. (2005). Hydrochory increases riparian plant species richness: A comparison between a free-flowing and a regulated river. *Journal of Ecology*, 93(6), 1094– 1103. https://doi.org/10.1111/j.1365-2745.2005.01057.x
- Januchowski-Hartley, S. R., McIntyre, P. B., Diebel, M., Doran, P. J., Infante, D. M., Joseph, C., & Allan, J. D. (2013). Restoring aquatic

- ecosystem connectivity requires expanding inventories of both dams and road crossings. Frontiers in Ecology and the Environment, 11(4), 211–217. https://doi.org/10.1890/120168
- Johansson, M., & Nilsson, C. (1993). Hydrochory, population dynamics and distribution of the clonal aquatic plant *Ranunculus lingua*. *Journal* of Ecology, 81(1), 81–91. https://doi.org/10.2307/2261226
- Johnson, L. E., Ricciardi, A., & Carlton, J. T. (2001). Overland dispersal of aquatic invasive species: A risk assessment of transient recreational boating. *Ecological Applications*, 11(6), 1789–1799. https://doi. org/10.1890/1051-0761(2001)011[1789:ODOAIS]2.0.CO;2
- Johnson, P. T. L., Olden, J. D., & Vander Zanden, M. J. (2008). Dam invaders: Impoundments facilitate biological invasions of freshwaters. Frontiers in Ecology and the Environment, 6, 357–363.
- Jones, J., Börger, L., Tummers, J., Jones, P., Lucas, M., Kerr, J., ... Garcia de Leaniz, C. (2019). A comprehensive assessment of stream fragmentation in Great Britain. Science of the Total Environment, 673, 756–762. https://doi.org/10.1016/j.scitotenv.2019.04.125
- Jones, J. J., Collins, A. L., Naden, P., & Sear, D. A. (2012). The relationship between fine sediment and macrophytes in rivers. *River Research and Applications*, 28, 1006–1018. https://doi.org/10.1002/rra.1486
- Kelly, N. E., Wantola, K., Weisz, E., & Yan, N. D. (2013). Recreational boats as a vector of secondary spread for aquatic invasive species and native crustacean zooplankton. *Biological Invasions*, 15(3), 509–519. https://doi.org/10.1007/s10530-012-0303-0
- Landolt, E. (1986). Biosystematic investigations in the family of duck-weeds (Lemnaceae) (vol. 2), the family of Lemnaceae—a monographic study (vol. 1). Veröffentlichungen des Geobotanischen Institutes Der Eidg. Techniasche Hochschule, Stiftung Rubel, Zürich.
- Langerhans, R. B., & Reznick, D. N. (2010). Ecology and evolution of swimming performance in fishes: Predicting evolution with biomechanics. In P. Domenici & B. G. Kap (Eds.), Fish locomotion: An eco-ethological perspective (pp. 220-248). Enfield, NH: Science Publishers.
- Lehner, B., Liermann, C. R., Revenga, C., Vörösmarty, C., Fekete, B., Crouzet, P., ... Wisser, D. (2011). High-resolution mapping of the world's reservoirs and dams for sustainable river-flow management. Frontiers in Ecology and the Environment, 9(9), 494–502. https://doi. org/10.1890/100125
- Lucas, M. C., & Baras, E. (2001). Migration of freshwater fishes. Oxford, UK: Blackwell Science.
- Lucas, M. C., & Batley, E. (1996). Seasonal movements and behaviour of adult barbel *Barbus barbus*, a riverine cyprinid fish: Implications for river management. *Journal of Applied Ecology*, 1345–1358. https:// doi.org/10.2307/2404775
- Madsen, J. D., Eichler, L. W., & Boylen, C. W. (1988). Vegetative spread of Eurasian watermilfoil in Lake George, New York. *Journal of Aquatic Plant Management*, 26, 47–50.
- Magilligan, F. J., & Nislow, K. H. (2005). Changes in hydrologic regime by dams. Geomorphology, 71(1-2), 61-78. https://doi.org/10.1016/j. geomorph.2004.08.017
- Makrakis, M. C., Miranda, L. E., Makrakis, S., Xavier, A. M. M., Fontes, H. M., & Morlis, W. G. (2007). Migratory movements of pacu, *Piaractus mesopotamicus*, in the highly impounded Parana River. *Journal of Applied Ichthyololgy*, 23, 700–704.
- Markwith, S. H., & Leigh, D. S. (2008). Subaqueous hydrochory: Open-channel hydraulic modelling of non-buoyant seed movement. Freshwater Biology, 53(11), 2274–2286. https://doi.org/10.1111/j.1365-2427.2008.02054.x
- Markwith, S. H., & Leigh, D. S. (2012). Comparison of estimated and experimental subaqueous seed transport. *Ecohydrology*, 5(3), 346–350. https://doi.org/10.1002/eco.225
- Markwith, S. H., & Scanlon, M. J. (2007). Multiscale analysis of Hymenocallis coronaria (Amaryllidaceae) genetic diversity, genetic structure, and gene movement under the influence of unidirectional stream flow. American Journal of Botany, 94(2), 151–160.

- Medwecka-Kornaś, A., & Hawro, R. (1993). Vegetation on beaver dams in the Ojcow National Park (Southern Poland). *Phytocoenologia*, 23, 611–618.
- Merritt, D. M., Nilsson, C., & Jansson, R. (2010). Consequences of propagule dispersal and river fragmentation for riparian plant community diversity and turnover. *Ecological Monographs*, 80, 609–626.
- Merritt, D. M., & Wohl, E. E. (2002). Processes governing hydrochory along rivers: Hydraulics, hydrology, and dispersal phenology. *Ecological Applications*, 12, 1071–1087.
- Merritt, D. M., & Wohl, E. E. (2006). Plant dispersal along rivers fragmented by dams. River Research and Applications, 22(1), 1–26. https://doi.org/10.1002/rra.890
- Michelan, T. S., Thomaz, S. M., Mormul, R. P., & Carvalho, P. (2010). Effects of an exotic invasive macrophyte (tropical signalgrass) on native plant community composition, species richness and functional diversity. *Freshwater Biology*, 55(6), 1315–1326. https://doi. org/10.1111/j.1365-2427.2009.02355.x
- Morita, K., & Yamamoto, S. (2002). Effects of habitat fragmentation by damming on the persistence of stream-dwelling charr populations. *Conservation Biology*, 16(5), 1318–1323. https://doi.org/10.1046/i.1523-1739.2002.01476.x
- Mueller, M., Pander, J., & Geist, J. (2011). The effects of weirs on structural stream habitat and biological communities. *Journal of Applied Ecology*, 48(6), 1450–1461. https://doi.org/10.1111/j.1365-2664.2011.02035.x
- Nathan, R., Schurr, F. M., Spiegel, O., Steinitz, O., Trakhtenbrot, A., & Tsoar, A. (2008). Mechanisms of long-distance seed dispersal. *Trends in Ecology & Evolution*, 23(11), 638–647. https://doi.org/10.1016/j.tree.2008.08.003
- Nilsson, C., Brown, R. L., Jansson, R., & Merritt, D. M. (2010). The role of hydrochory in structuring riparian and wetland vegetation. *Biological Reviews*, 85, 837–858.
- Nilsson, C., & Dynesius, M. (1994). Ecological effects of river regulation on mammals and birds: A review. Regulated Rivers: Research & Management, 9(1), 45-53. https://doi.org/10.1002/rrr.34500 90105
- Nilsson, C., Ekblad, A., Gardfjell, M., & Carlberg, B. (1991). Long-term effects of river regulation on river-margin vegetation. *Journal of Applied Ecology*, 28, 963–987.
- Nilsson, C., Gardfjell, M., & Grelsson, G. (1991). Importance of hydrochory in structuring plant communities along rivers. *Canadian Journal* of *Botany*, 69(12), 2631–2633. https://doi.org/10.1139/b91-328
- Nilsson, C., & Jansson, R. (1995). Floristic differences between riparian corridors of regulated and free-flowing boreal rivers. *Regulated Rivers: Research and Management*, 11, 55-66.
- Nurminen, L., Horppila, J., Lappalainen, J., & Malinen, T. (2003). Implications of rudd (*Scardinius erythrophthalmus*) herbivory on submerged macrophytes in a shallow eutrophic lake. *Hydrobiologia*, 506–509, 511–518. https://doi.org/10.1023/B:HYDR.0000008577.16934.a9
- Okada, M., Grewell, B. J., & Jasieniuk, M. (2009). Clonal spread of invasive *Ludwigia hexapetala* and *L. grandiflora* in freshwater wetlands of California. *Aquatic Botany*, 91, 123–129. https://doi.org/10.1016/j.aquabot.2009.03.006
- Olden, J. D., & Naiman, R. J. (2010). Incorporating thermal regimes into environmental flows assessments: Modifying dam operations to restore freshwater ecosystem integrity. *Freshwater Biology*, 55(1), 86–107. https://doi.org/10.1111/j.1365-2427.2009.02179.x
- Padgett, D. J., Carboni, J. J., & Schepis, D. J. (2010). The dietary composition of *Chrysemys picta picta* (eastern painted turtles) with special reference to the seeds of aquatic macrophytes. *Northeastern Naturalist*, 17(2), 305–313.
- Parasiewicz, P., Schmutz, S., & Moog, O. (1998). The effect of managed hydropower peaking on the physical habitat, benthos and fish fauna in the River Bregenzerach in Austria. *Fisheries Management and Ecology*, 5, 403–417.

- Perkin, J. S., & Gido, K. B. (2012). Fragmentation alters stream fish community structure in dendritic ecological networks. *Ecological Applications*, 22(8), 2176–2187. https://doi.org/10.1890/12-0318.1
- Pollux, B. J. A. (2007). Plant dispersal in rivers A mechanistic and molecular approach. PhD thesis, Radboud. University Nijmegen, Nijmegen.
- Pollux, B. J. A. (2011). The experimental study of seed dispersal by fish (ichthyochory). *Freshwater Biology*, 56(2), 197–212. https://doi.org/10.1111/j.1365-2427.2010.02493.x
- Pollux, B. J. A., De Jong, M., Steegh, A., Ouborg, N. J., Van Groenendael, J. M., & Klaassen, M. (2006). The effect of seed morphology on the potential dispersal of aquatic macrophytes by the common carp (Cyprinus carpio). Freshwater Biology, 51(11), 2063–2071. https://doi.org/10.1111/j.1365-2427.2006.01637.x
- Pollux, B. J. A., Luteijn, A., Van Groenendael, J. M., & Ouborg, N. J. (2009). Gene flow and genetic structure of the aquatic macrophyte Sparganium emersum in a linear unidirectional river. Freshwater Biology, 54(1), 64–76.
- Pollux, B. J. A., Ouborg, N. J., van Groenendael, J. M., & Klaassen, M. (2007). Consequences of intraspecific seed-size variation in *Sparganium emersum* for dispersal by fish. *Functional Ecology*, 21, 1084–1091. https://doi.org/10.1111/j.1365-2435.2007.01313.x
- Pollux, B. J. A., Santamaria, L., & Ouborg, N. J. (2005). Differences in endozoochorous dispersal between aquatic plant species, with reference to plant population persistence in rivers. *Freshwater Biology*, 50(2), 232–242. https://doi.org/10.1111/j.1365-2427.2004.01314.x
- Pringle, C. M. (2001). Hydrologic connectivity and the management of biological reserves: A global perspective. *Ecological Applications*, 11(4), 981–998. https://doi.org/10.1890/1051-0761(2001)011[0981:H-CATMO]2.0.CO;2
- Pringle, C. M. (2003). What is hydrologic connectivity and why is it ecologically important? *Hydrological Processes*, 17(13), 2685–2689. https://doi.org/10.1002/hyp.5145
- Reid, A. J., Carlson, A. K., Creed, I. F., Eliason, E. J., Gell, P. A., Johnson, P. T. J., ... Cooke, S. J. (2019). Emerging threats and persistent conservation challenges for freshwater biodiversity. *Biological Reviews*, 94(3), 849–873. https://doi.org/10.1111/brv.12480
- Reynolds, C., & Cumming, G. S. (2016). Seed dispersal by waterbirds in southern Africa: Comparing the roles of ectozoochory and endozoochory. Freshwater Biology, 61(4), 349–361. https://doi.org/10.1111/fwb.12709
- Riis, T. (2008). Dispersal and colonisation of plants in lowland streams: Success rates and bottlenecks. *Hydrobiologia*, *596*(1), 341–351. https://doi.org/10.1007/s10750-007-9107-0
- Riis, T., & Biggs, B. J. F. (2003). Hydrologic and hydraulic control of macrophyte establishment and performance in streams. *Limnology* and Oceanography, 48, 1488–1497. https://doi.org/10.4319/ lo.2003.48.4.1488
- Riis, T., Madsen, T. V., & Sennels, R. S. H. (2009). Regeneration, colonisation and growth rates of allofragments in four common stream plants. *Aquatic Botany*, *90*(2), 209–212. https://doi.org/10.1016/j. aquabot.2008.08.005
- Riis, T., & Sand-Jensen, K. (2006). Dispersal of plant fragments in small streams. *Freshwater Biology*, *5*1(2), 274–286. https://doi.org/10.1111/j.1365-2427.2005.01496.x
- Rood, S. B., Braatne, J. H., & Goater, L. A. (2010). Favorable fragmentation: River reservoirs can impede downstream expansion of riparian weeds. *Ecological Applications*, 20(6), 1664–1677. https://doi.org/10.1890/09-0063.1
- Rorslett, B., & Johansen, S. W. (1996). Remedial measures connected with aquatic macrophytes in Norwegian regulated rivers and reservoirs. Regulated Rivers-Research & Management, 15, 509–522. https:// doi.org/10.1002/(SICI)1099-1646(199607)12:4/5<509:AID-RRR41 0>3.0.CO;2-3
- Rothlisberger, J. D., Chadderton, W. L., McNulty, J., & Lodge, D. M. (2010). Aquatic invasive species transport via trailered boats: What is being moved, who is moving it, and what can be done. *Fisheries*, *35*(3), 121–132. https://doi.org/10.1577/1548-8446-35.3.121

Sand-Jensen, K., Andersen, K., & Andersen, T. (1999). Dynamic properties of recruitment, expansion and mortality of macrophyte patches in streams. International Review of Hydrobiology, 84(5), 497-508.

- Sand-Jensen, K. A. J., Jeppesen, E., Nielsen, K., Van der Bijl, L., Hjermind, L., Nielsen, L. W., & IvrlsIn, T. M. (1989). Growth of macrophytes and ecosystem consequences in a lowland Danish stream. Freshwater Biology, 22(1), 15-32. https://doi.org/10.1111/j.1365-2427.1989.tb01080.x
- Saura, S., Bodin, Ö., & Fortin, M. J. (2014). EDITOR'S CHOICE: Stepping stones are crucial for species' long-distance dispersal and range expansion through habitat networks. Journal of Applied Ecology, 51(1), 171-182. https://doi.org/10.1111/1365-2664.12179
- Schultz, R., & Dibble, E. (2012). Effects of invasive macrophytes on freshwater fish and macroinvertebrate communities: The role of invasive plant traits. Hydrobiologia, 684(1), 1-14. https://doi.org/10.1007/ s10750-011-0978-8
- Shipley, B., Keddy, P. A., Moore, D. R. J., & Lemky, K. (1989). Regeneration and establishment strategies of emergent macrophytes. Journal of Ecology, 77, 1093-1110.
- Silva, G. G., Green, A. J., Weber, V., Hoffmann, P., Lovas-Kiss, Á., Stenert, C., & Maltchik, L. (2018). Whole angiosperms Wolffia columbiana disperse by gut passage through wildfowl in South America. Biology Letters, 14(12), 20180703.
- Skoglund, S. J. (1989). Seed dispersal agents in two regularly flooded river sites. Canadian Journal of Botany, 68, 754-760.
- Smits, A. J. M., Van Ruremonde, R., & Van Der Velde, G. (1989). Seed dispersal of three nymphaeid macrophytes. Aquatic Botany, 35(2), 167-180. https://doi.org/10.1016/0304-3770(89)90103-4
- Soomers, H., Karssenberg, D., Soons, M. B., Verweij, P. A., Verhoeven, J. T., & Wassen, M. J. (2013). Wind and water dispersal of wetland plants across fragmented landscapes. Ecosystems, 16(3), 434-451. https://doi.org/10.1007/s10021-012-9619-y
- Soons, M. B. (2006). Wind dispersal in freshwater wetlands: Knowledge for conservation and restoration. Applied Vegetation Science, 9(2), 271-278. https://doi.org/10.1111/j.1654-109X.2006.tb00676.x
- Staniforth, R. J., & Cavers, P. B. (1976). An experimental study of water dispersal in Polygonum spp. Canadian Journal of Botany, 54, 2587-2596.
- Stevens, L. E., Buck, K. A., Brown, B. T., & Kline, N. C. (1997). Dam and geomorphological influences on Colorado River waterbird distribution, Grand Canyon, Arizona, USA. Regulated Rivers: Research & Management, 13(2), 151-169. https://doi.org/10.1002/(SICI)1099-1646(199703)13:2<151:AID-RRR447>3.0.CO;2-U
- Strayer, D. L. (2010). Alien species in fresh waters: Ecological effects, interactions with other stressors, and prospects for the future. Freshwater Biology, 55, 152-174. https://doi. org/10.1111/j.1365-2427.2009.02380.x
- Tamayo, M., & Olden, J. D. (2014). Forecasting the vulnerability of lakes to aquatic plant invasions. Invasive Plant Science and Management, 7(1), 32-45. https://doi.org/10.1614/IPSM-D-13-00036.1
- Téllez, T. R., López, E. M. D. R., Granado, G. L., Pérez, E. A., López, R. M., & Guzmán, J. M. S. (2008). The water hyacinth, Eichhornia crassipes: An invasive plant in the Guadiana River Basin (Spain). Aquatic Invasions, 3(1), 42-53. https://doi.org/10.3391/ai.2008.3.1.8
- Tena, A., Vericat, D., Gonzalo, L. E., & Batalla, R. J. (2017). Spatial and temporal dynamics of macrophyte cover in a large regulated river. Journal of Environmental Management, 202, 379-391. https://doi. org/10.1016/j.jenvman.2016.11.034
- Tero, N., Aspi, J., Siikamäki, P., Jäkäläniemi, A., & Tuomi, J. (2003). Genetic structure and gene flow in a metapopulation of an endangered plant species, Silene tatarica. Molecular Ecology, 12(8), 2073-2085. https:// doi.org/10.1046/j.1365-294X.2003.01898.x
- Thébaud, C., & Debussche, M. (1991). Rapid invasion of Fraxinus ornus L. along the Herault River System in southern France: The importance

- of seed dispersal by water. Journal of Biogeography, 18, 7-12. https:// doi.org/10.2307/2845240
- Thomason, M. J. S., McCort, C. D., Netherland, M. D., & Grewell, B. J. (2018). Temporal and nonlinear dispersal patterns of Ludwigia hexapetala in a regulated river. Wetlands Ecology and Management, 26(5), 751-762. https://doi.org/10.1007/s11273-018-9605-z
- Thomaz, S. M., Mormul, R. P., & Michelan, T. S. (2015). Propagule pressure, invasibility of freshwater ecosystems by macrophytes and their ecological impacts: A review of tropical freshwater ecosystems. Hydrobiologia, 746(1), 39-59. https://doi.org/10.1007/ s10750-014-2044-9
- Tombolini, I., Caneva, G., Cancellieri, L., Abati, S., & Ceschin, S. (2014). Damming effects on upstream riparian and aquatic vegetation: The case study of Nazzano (Tiber River, central Italy). Knowledge and Management of Aquatic Ecosystems, 412(3), 1-15. https://doi. org/10.1051/kmae/2013085
- Trakhtenbrot, A., Nathan, R., Perry, G., & Richardson, D. M. (2005). The importance of long-distance dispersal in biodiversity conservation. Diversity and Distributions, 11(2), 173-181. https://doi. org/10.1111/j.1366-9516.2005.00156.x
- Tudorache, C., Viaene, P., Blust, R., Vereecken, H., & De Boeck, G. (2008). A comparison of swimming capacity and energy use in seven European freshwater fish species. Ecology of Freshwater Fish, 17(2), 284-291. https://doi.org/10.1111/j.1600-0633.2007.00280.x
- Umetsu, C. A., Evangelista, H. B. A., & Thomaz, S. M. (2012). The colonization, regeneration, and growth rates of macrophytes from fragments: A comparison between exotic and native submerged aquatic species. Aquatic Ecology, 46(4), 443-449. https://doi.org/10.1007/ s10452-012-9413-0
- Van Geest, G. J., Coops, H., Roijackers, R. M. M., Buijse, A. D., & Scheffer, M. (2005). Succession of aquatic vegetation driven by reduced water-level fluctuations in floodplain lakes. Journal of Applied Ecology, 42(2), 251-260. https://doi. org/10.1111/j.1365-2664.2005.00995.x
- Van Leeuwen, C. H., Lovas-Kiss, Á., Ovegård, M., & Green, A. J. (2017). Great cormorants reveal overlooked secondary dispersal of plants and invertebrates by piscivorous waterbirds. Biology Letters, 13(10), 20170406. https://doi.org/10.1098/rsbl.2017.0406
- Van Leeuwen, C. H. A., van der Velde, G., van Groenendael, J. M., & Klaassen, M. (2012). Gut travellers: Internal dispersal of aquatic organisms by waterfowl. Journal of Biogeography, 39(11), 2031-2040. https://doi.org/10.1111/jbi.12004
- VonBank, J. A., Casper, A. F., Pendleton, J. E., & Hagy, H. M. (2018). Water hyacinth (Eichhornia crassipes) invasion and establishment in a temperate river system. River Research and Applications, 34(10), 1237-1243.
- VonBank, J. A., DeBoer, J. A., Casper, A. F., & Hagy, H. M. (2018). Ichthyochory in a temperate river system by common carp (Cyprinus carpio). Journal of Freshwater Ecology, 33(1), 83-96.
- Vukov, D., Ilić, M., Ćuk, M., Radulović, S., Igić, R., & Janauer, G. A. (2018). Combined effects of physical environmental conditions and anthropogenic alterations are associated with macrophyte habitat fragmentation in rivers-Study of the Danube in Serbia. Science of the Total Environment, 634, 780-790. https://doi.org/10.1016/j.scito tenv.2018.03.367
- Ward, J. V., & Stanford, J. A. (1983). The serial discontinuity concept of lotic ecosystems. In T. D. Fontaine & S. M. Bartell (Eds.), Dynamics of lotic ecosystems (pp. 29-42). Ann Arbor, MI: Ann Arbor Scientific Publishers.
- Ward, J. V., & Stanford, J. A. (1995). Ecological connectivity in alluvial river ecosystems and its disruption by flow regulation. Regulated Rivers: Research & Management, 11(1), 105-119. https://doi.org/10.1002/ rrr.3450110109
- Welham, C. V. J. (1994). Flight speeds of migrating birds: A test of maximum range speed predictions from the aerodynamic equations. Behavioural Ecology, 5, 1-8.

- Wichmann, M. C., Alexander, M. J., Soons, M. B., Galsworthy, S., Dunne, L., Gould, R., ... Bullock, J. M. (2008). Human-mediated dispersal of seeds over long distances. *Proceedings of the Royal Society B: Biological Sciences*, 276(1656), 523–532.
- Williams, G. P., & Wolman, M. G. (1984). Downstream effects of dams on alluvial rivers. United States Geological Survey Professional Paper 1286
- Winter, H. V., & Van Densen, W. L. T. (2001). Assessing the opportunities for upstream migration of non-salmonid fishes in the weir-regulated River Vecht. *Fisheries Management and Ecology*, 8(6), 513–532. https://doi.org/10.1046/j.1365-2400.2001.00271.x
- Wright, S. (1931). Evolution in Mendelian populations. *Genetics*, 16, 114–138.

# SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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